

**VERIFICATION OF NON-ORGANIC INGREDIENT STATUS**

*This form must be completed by the supplier or manufacturer of a non-organic ingredient used by PCO-certified operations in products labeled as “organic” or “made with organic (specified food groups).”. PCO may require additional information if needed to verify compliance with applicable regulations and policies. Relevant definitions are included on page 2.*

The USDA National Organic Program regulations allow for the use of non-organic ingredients that are produced and handled without the use of excluded methods, ionizing radiation, and sewage sludge (7 CFR 205.105(e)-(g)).

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| **Ingredient:** |  |
| **Supplier/Manufacturer Name:** |  |

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| 1. The ingredient listed above is produced and handled without the use of excluded methods, genetic engineering, or genetically manipulated organisms or ingredients, as described on page 2. This product is not derived from products or ingredients that contain genetically modified organisms (GMO) and has not been produced with GMO processing aids. *Microbial substrate, feedstocks, or culture media consumed or removed are not required to be produced without excluded methods.* | True | False | |
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| 1. The ingredient listed above is produced and handled without the use of Ionizing radiation, as described in the FDA regulation 21 CFR 179.26. | True | False | |
|  |  |  | |
| 1. This ingredient is produced and handled without the use of sewage sludge. | True | False | |
|  |  |  | |
| 1. Check here to OMIT this material from PCO’s printed materials list (*e.g. custom or proprietary inputs).* | OMIT | | |
|  |  | | |
| 1. *For yeast:* This yeast is not grown on petrochemical substrate or sulfite waste liquor. | True | False | N/A |
|  |  |  |  |
| 1. *For citric acid products:* This citric acid is produced by microbial fermentation of a carbohydrate substance. | True | False | N/A |
|  |  |  |  |
| 1. *For enzymes:* This enzyme is derived from edible, nontoxic plants, nonpathogenic fungi, or nonpathogenic bacteria. | True | False | N/A |

*To be signed by the manufacturer or supplier. Signer must be a qualified technical person.*

**Pursuant to applicable regulations, I, on behalf of the supplier or manufacturer, hereby attest that the information provided in this form is accurate and truthful to the best of my knowledge.**

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| --- | --- | --- |
| Signature: | | Date: |
| Printed Name: | Title: | |
| Address: | | |
| City: | State: | Zip: |
| Phone: | Email: | |

**Excluded methods** are defined at 7 CFR 205.2 as a variety of methods used to genetically modify organisms or influence their growth and development by means that are not possible under natural conditions or processes and are not considered compatible with organic production. Such methods include cell fusion, microencapsulation and macroencapsulation, and recombinant DNA technology (including gene deletion, gene doubling, introducing a foreign gene, and changing the positions of genes when achieved by recombinant DNA technology). Such methods do not include the use of traditional breeding, conjugation, fermentation, hybridization, in vitro fertilization, or tissue culture.

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| ***Method and synonyms*** | ***Types*** |
| Targeted genetic modification (TagMo)  syn. Synthetic gene technologies  syn. Genome engineering  syn. Gene editing  syn. Gene targeting | Sequence-specific nucleases (SSNs)  Meganucleases Zinc finger nuclease (ZFN)  Mutagenesis via Oligonucleotides  CRISPR-Cas system (Clustered regularly interspaced short palindromic repeats) and associated protein genes  TALENs (Transcription activator-like effector nucleases)  Oligonucleotide directed mutagenesis (ODM) Rapid Trait Development System |
| Gene Silencing | RNA-dependent DNA methylation (RdDM) Silencing via RNAi pathway RNAi pesticides |
| Accelerated plant breeding techniques | Reverse Breeding  Genome Elimination  FasTrack  Fast flowering |
| Synthetic biology | Creating new DNA sequences  Synthetic chromosomes  Engineered biological functions and systems |
| Cloned animals and offspring | Somatic nuclear transfer |
| Plastic transformation |  |
| Cisgenesis | The gene modification of a recipient plant with a natural gene from a crossable-sexually compatible-plant. The introduced gene includes its introns and is flanked by its native promoter and terminator in the normal-sense orientation. |
| Intragenesis | The full or partial coding of DNA sequences of genes originating from the sexually compatible gene pool of the recipient plant and arranged in sense or antisense orientation. In addition, the promoter, spacer, and terminator may originate from a sexually compatible gene pool of the recipient plant. |
| Agro-infiltration |  |
| Transposons – Developed via use of in vitro nucleic acid techniques |  |
| Induced mutagenesis | Developed through in vitro nucleic acid techniques |